Helminth Fauna of *Oreochromis leucostictus* (Pisces: CICHLIDAE) from a tropical lake, Lake Naivasha, Kenya

*P.A. ALOO1, R.O. OKELO1, AND M.J. NTIBA2

- Department of Zoology
 Kenyatta University
 P.O. Box 51336, Nairobi, Kenya
- Department of Zoology
 University of Nairobi
 P.O. Box 30197, Nairobi, Kenya
- * Corresponding Author

ABSTRACT

The helminth fauna of *Oreochromis leucostictus* (Trewavas) from Lake Naivasha is described. The study was carried out on 1,050 gillnetted fish caught over a period of 24 months at different sites within the lake. *Oreochromis leucostictus* which is a commercially important fish was observed to be a host of four helminth parasites. These were: an acanthocephalan *Polyacanthorhynchus kenyensis* (Schmidt and Canaris), a nematode *Contracaecum* sp., a trematode *Clinostomum* sp. and a dilepidid cestode *Amirthalingamia* sp.

Seasonal variation in prevalence, intensity of infection, organs infected and the variation in infection with the size, sex and body condition of the host are described. There was no seasonal variation in prevalence but intensity of infection was observed to increase with the age of the fish. Although male fish were observed to be more heavily infected, the parasites did not seem to have any significant effect on the body condition of the fish.

INTRODUCTION

Lake Naivasha is the only freshwater lake in the eastern arm of the Kenyan portion of the Rift Valley. It is situated about 100 km north of Nairobi, the capital city (Fig. 1). The fish fauna of this lake is made up of three introduced teleostean species; Oreochromis leucostictus (Trewavas), Micropterus salmoides (Lace'p'ede) and Tilapia zillii (Gervais). These three species form the mainstay of a commercial fishery, which has been established in the lake for over 40 years - with O. leucostictus being the most important commercial fish. Also present in Lake Naivasha are the riverine species, Barbus amphigramma (Boulenger), and the Red swamp crayfish, Procambarus clarkii (Girrard). The various aspects of the ecology of these fish species have received adequate attention, but the only reports available on the parasitic fauna are those of MALVESTUTO (1975)Contracaecum sp. of O. leucostictus and MALVESTUTO and OGAMBO-ONGOMA (1978) on the infection rate of O. leucostictus with the same parasite. The present study reports on the occurrence of other parasites in this fish.

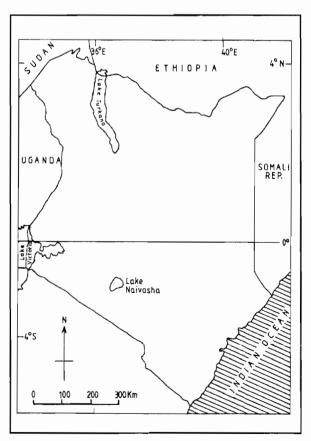


Fig. 1. A map of Kenva showing the location of Lake Naivasha

MATERIALS AND METHODS

Fish were caught from 13 different sampling stations established around the lake (Fig. 2) between October 1991 and June 1994. The stations were selected based on broad categories of different habitats found in the lake, (e.g. rocky shores, papyrus fringes and submerged macrophyte beds). A fleet of 8 gillnets of mesh sizes 1¹/2, 2", 2¹/2, 3", 3¹/2, 4", 4¹/2 and 5" were laid at each station overnight starting from 6 a.m. to 10 a.m. and the fish collected the following day at the same time. The total number of *O. leucostictus* caught at each station was enumerated to establish its abundance in the different habitats.

During removal from the net, fish were thoroughly examined to recover parasites such as monogeneans that may fall off after removal of the fish from water. Fish samples were then transported to the laboratory by the shores of the lake. In the laboratory, the lengths of fish from each station were measured and the fish divided into 5 cm size classes. Sex of each fish was also recorded and subsamples from each station was drawn based on sex and size. For ectoparasites, the fish were placed in containers under running water, scrubbed using a brush, and the washings sieved and examined under a dissecting microscope. Various organs such as around the fins, beneath the operculum, nostrils and eyes were examined for ectoparasites.

Endoparasites were recovered by opening the fish dorso-ventrally to expose the internal organs. The entire digestive system was removed and placed in a petri-dish containing saline solution. The stomach and the intestines were separated and each opened up and searched for parasites. Other internal organs examined for parasites included the gonads and urinary bladder.

The parasites found were subjected to different treatments as follows: Nematodes were killed in 70% alcohol which made them remain stretched for easy counting and taking morphometric measurements. Acanthocephalans and cestodes were placed in petri-dishes of cold water and kept in a refrigerator overnight to facilitate the extrusion of the proboscis and scolex principally used as taxonomical features in their identification. Trematodes were pressed

between 2 slides with a drop of glacial acetic acid which made them transparent so that internal organs were visible under a dissecting microscope. Permanent whole mount preparations were then made for taxonomical studies. The effect of Contracaecum and P. kenyensis on the health status of O. leucostictus was investigated by establishing the relationship between parasite intensity and the condition factor (K-factor) of the fish and K-factor was calculated using the formula:

$$K = \frac{L^3}{100W}$$
 where: $L = length (cm)$
 $W = weight (g)$

Statistical analysis used during this work included Split-Plot technique, regression analysis and Chi-square tests. The terms prevalence and intensity are used in this paper as defined by MARGOLIS *et al.*, (1982).

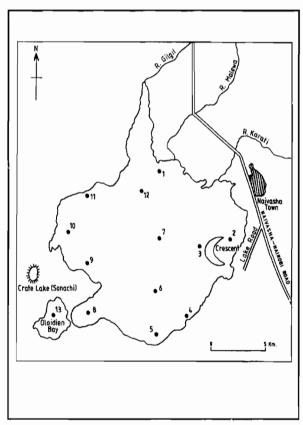


Fig. 2. A map of Lake Naivasha showing Sampling Stations

Stations

- 1. North Swamp
- 2. Crescent Island
- 3. Flamingo Farm
- 4. Safariland
- 5. Fisherman's Camp
- 6. Lakeside Farm
- 7. Middle Lake
- 8. Oseria
- 9. Hippo Point
- 10. White House
- 11. Korongo
- 12. Hop Craft
- 13. Oloidien

RESULTS

Oreochromis leucostictus from Lake Naivasha did not harbour any ectoparasites. However, the fish was a host to four helminth parasites which were abundant in the following order: cystacanths of *Polyacanthorhynchus kenyensis* (Schmidt and Canaris) found encysted in the

liver; third stage larvae of a nematode, Contracaecum sp., which were found free in the pericardial cavity; metacercariae of a trematode Clinostomum sp. which occurred in the pharyngeal region; and cysticercoids of a dilepidid cestode Amirithalingamia sp. which were either encysted in the liver or free floating in the intestines (Figures 3 - 6).

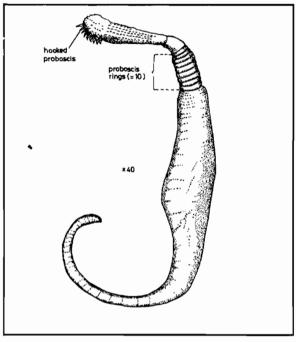


Fig. 3. Polyacanthorbynchus kenyensis

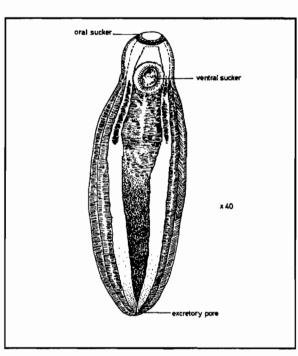


Fig. 5. Clinostomum sp.

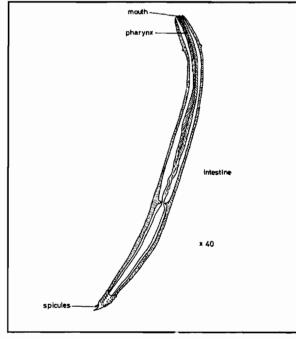


Fig. 4. Contracaecum sp.

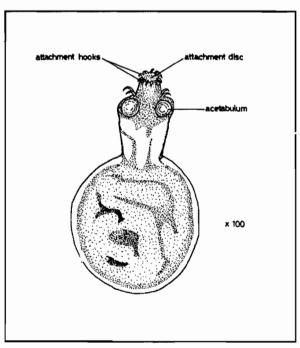


Fig. 6. Amirthaligamia sp. (Cystercercoid)

The parasites did not show any seasonality in the intensity of infection throughout the study period. The results also showed no significant variation in prevalence between months (Fig. 7).

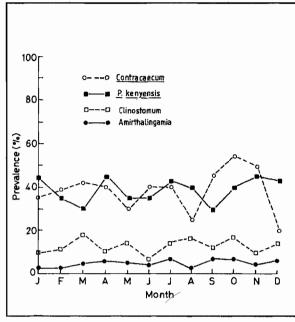


Fig. 7. Monthly Variation in the Prevalence of Parasites in Oreochromis leucostictus in Lake Naivasha

However, a very significant variation was observed in the prevalence of the Contracaecum within sampling stations, with Oloidien bay recording the highest number of infected hosts while towards the middle of the lake no infected hosts were found (Table I). Another significant variation was observed in the intensity of infection at various sites where O. leucostictus from Oloidien were more heavily infected by the nematode Contracaecum than fish caught at other stations (Table I) (maximum intensity = 68parasites per host). Polyacanthorynchus kenyensis was observed to occur more abundantly in hosts caught at North Swamp (max. intensity = 104 parasites), Korongo (max. intensity = 98 parasites) and Cresent (max. intensity = 52 parasites); while Amirthalingamia sp. were observed to infect mainly fish from Oloidien (F_{60} , 2,860 = 7.79; P<0.001).

Regression analysis revealed that there was non-significant inverse relationship between the intensity of infection with *Contracaecum* and the size of the fish. However, the intensity of infection with *P. kenyensis* increased significantly with increase in the size of the fish $(t_4 = 7.06; P<0.001)$ (Tables II and III).

Highly significant relationships were observed between the prevalence of *Contracaecum* and *P. kenyensis* and the size of *O. leucostictus*. This was more evident in the case of *P. kenyensis* ($t_4 = 3.8$; P<0.002) (Fig. 8).

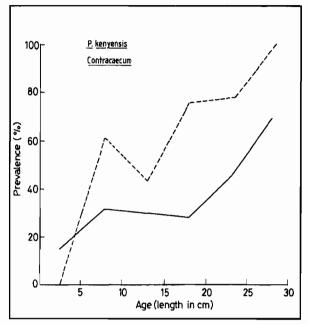


Fig. 8. Prevalence of *Contracaecum* and *P. kenyensis* with the Age of *O. leucostictus*

Using Chi-square analysis, it was observed that there was a very significant relationship between the sex of O. leucostictus and the intensity of infection ($X^2_{(1)} = 64.43$; P<0.001). Male fish were observed to harbour more parasites than their female counterparts (Contracaecum = 205 in females vs 908 in males; and P. kenyensis = 778 in females vs 1766 in males) (n=540).

Results obtained have revealed that there was no significant relationship between K-factor and the number of worms (*Contracaecum* and *P. kenyensis*) (Fig. 9).

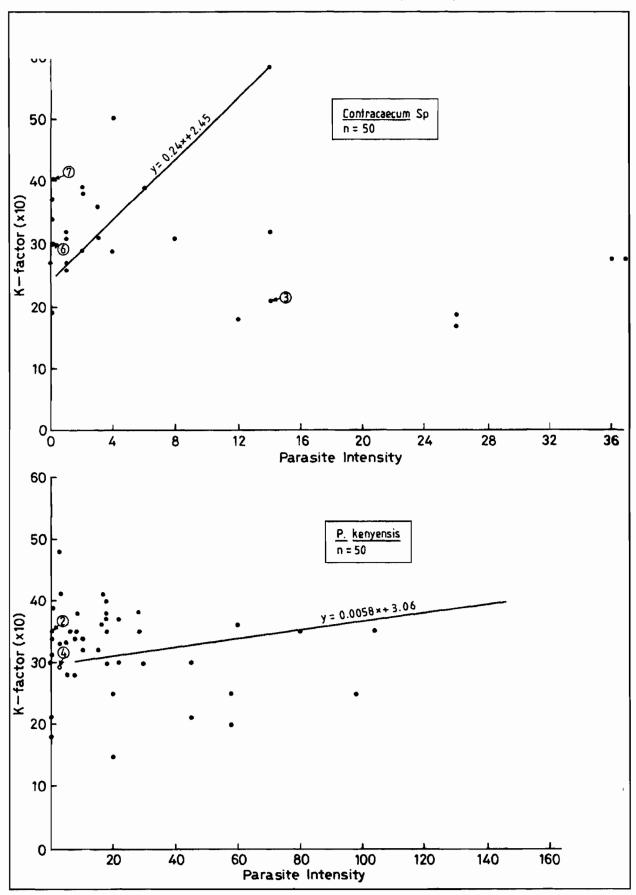


Fig. 9. The effect of the parasites on the health status of O. leocostictus

Table I Variation in total number of parasites within sampling stations

Total No. of Fish examined	165	180	80	9	70	58	0 (0)	69	105	74	81	42	61
Total No. of Armithalingamia	4	0	12 (8)	15.(7)	13 (6)	6 (4)	0 (0)	10 (6)	18 (11)	0 (0)	47 (16)	13 (10)	41 (24)
Total No. of Clinostomum	33 (22)	43 (20)	22 (18)	41 (27)	38 (19)	13 (11)	0 (0)	33 (14)	14 (10)	11 (7)	47 (23)	32 (12)	7 (5)
Total No. of P. kenyensis	577 (110)	526 (132)	156 (64)	348 (51)	347 (85)	110 (40)	0 (0)	345 (48)	478 (94)	132 (42)	488 (61)	141 (28)	90 (42)
Total No. of Contracaecum	39 (31)	163 (98)	19 (11)	75 (38)	139 (49)	24 (18)	0)0	43 (31)	27 (15)	38 (20)	54 (42)	32 (28)	1282 (61)
Stations	1. Northswamp	2. Crescent	3. Flamingo Farm	4. Flamingo	5. Fishermen's Camp	6. Lakeside Farm	7. Middle of lake	8. Oseria	9. Hippopoint	10. Whitehouse	11. Northswamp	12. Hopcraft	13. Oloidien

Numbers in parentheses indicates infected hosts

Table II Mean Intensity of infection of O. leucostictus with Contracaecum

Length class (cm)	No. of fish examined	No. of fish infected	Mean intensity (MI)	Total No. of parasites	Range of parasites
0 - 5	10	2	2.0	4	J passi
0 - 10	30	20	7.5	150	9 - 1
11 - 15	320	210	8.6	1800	1 - 14
16 - 20	362	172	1.4	1241	1 - 68
21 - 25	241	82	2.8	229	1 - 5
26 - 30	70	38	8.9	259	1 - 5
31 - 35	17	∞	1.9	15	1 - 2

Table III Mean Intensity of infection of O. leucostictus with P. kenyensis

Length class (cm)	No. of fish examined	No. of fish infected	Mean intensity (MI)	Total No. of parasites	Range of parasites
0 - 5	10	0	0	0	0
0 - 10	30	8	1.9	15	1-3
11 - 15	320	125	09	750	6 - 1
16 - 20	362	194	7.2	1,406	1 - 24
21 - 25	241	37	27.6	1,023	1 - 24
26 - 30	70	16	28.6	458	1 - 68
31 - 35	17	∞	10.8	98	1 - 104

DISCUSSION

Since the introduction of O. leucostictus in Lake Naivasha, the only probable explanation for the absence of ectoparasites is that these parasites are easily affected by any change in environmental factors such as salinity and temperature. Therefore, those that arrived into the lake with the fish may have disappeared due to the above changes. Various ectoparasites such as the argulids and ergasiliids have been reported to occur on the same fish in Lake Victoria from which O. leucostictus was introduced (FRYER and ILES, 1972, PAPERNA. 1980; 1991). PAPERNA (1980) also reported on the occurrence of monogenean trematodes on the tilapiine species of Lake Victoria. However, in Lake Naivasha even the other 3 fish species do not harbour any ectoparasites (ALOO, 1995). The present results have challenged those of MALVESTUTO and OGAMBO-ONGOMA (1978) and PAPERNA (1980) who reported that O. leucostictus from Lake Naivasha were infected only by Contracaecum sp.

The fact that there was no significant variation in intensity and prevalence between the months suggests that the fish are infected throughout the year. These findings do not agree with other reports from tropical waters in which the seasonality of fish parasites has been observed such as PAPERNA (1980) on Clinostomum sp. in tilapias in Kenya, MBAHINZIREKI (1980) on helminth parasites of Haplochromis spp. in Uganda and BATRA (1984) on helminth parasites of cichlids from Zambia. However, various reports on the acanthocephalans from temperature regions indicated that they infect their hosts throughout the year. These include the findings of SCHOLTZ (1986) on the Acanthocephalus lucii infecting Perca fluviatilis **AMIN** and (1985)Echinorhynchus salmonis infecting the rainbow smelt, Osmerus mordax. In the tropics there is no marked seasonality as in the temperature regions hence the occurrence of parasites throughout the year as was observed during this study. Since MALVESTUTO and OGAMBO-ONGOMA (1978) suggested that O. becomes infected leucostictus Contracaecum directly and not through an

intermediate host, it is therefore possible that the parasites will be found in the hosts throughout the year because seasonality will only have a direct effect on intermediate hosts. WILLIAM and JONES (1994) linked seasonality in prevalence of fish parasites to many ecological factors e.g. water temperatures, the feeding behaviour and the diet of the hosts.

Both prevalence and intensity of infection were observed to vary from one site to another. This was attributed to the differences in the physicochemical parameters among the sites. For example, the nematode Contracaecum was observed to be most abundant at Oloidien (site 13) which was a more saline environment, whereas the other parasites especially Clinostomum were present in very low numbers at the same site. These findings concur with those of DOGIEL et al., (1958) (In WILLIAM and JONES, 1994) that increase in salinity increases the prevalence and abundance of some parasites but also leads to the disappearance of those parasites whose intermediate host cannot withstand high salinity such as trematodes. The second point related to variation in prevalence and intensity from site to site is the size and isolation of site 13. Depletion of parasite fauna with isolation and size of habitats had been reported (DOGIEL et al, 1958) (In WILLIAM and JONES, 1994). This was attributed to the poor invertebrate fauna of small isolated water bodies: therefore parasites that require invertebrate intermediate hosts are predominated over by those with direct life cycles. The authors concluded that among the parasites observed, Contracaecum seem to have a direct life cycle hence their abundance in site 13.

The present study has shown a positive trend in both prevalence and intensity of infection by *Contracaecum* sp. and *P. kenyensis* with increase in the length of the fish. The results agree with a number of reports on the same subject such as PAPERNA (1980) who reported an increase in infection rate of tilapias by the trematode *Clinostomum* as the fish increased in size. MASHEGO (1989) also reported increased intensity of infection of *Barbus* sp by the nematode *Contracaecum* with increase in

fish length. Although the two did not give reasons for their observations, the present work indicates that in the case of Contracaecum it is possible the fish feeds more on the bottom materials containing larval worms, whereas in the case of P. kenyensis, the fish probably feed more on the supposed intermediate hosts (Ostracods) as they grow. Another explanation to this observation is that larval parasites will accumulate in the host over a period of time.

That male fish were observed to harbour more worms than their female counterparts is in accordance with the findings of other authors such as PALING (1965) who observed that male windermere trouts were more intensely infected than the females. He attributed this to a higher physiological resistance in females and to the large size of male fish which make them consume more food including the intermediate hosts hence more parasites. While working on the helminth parasites of the cichlids in Zambia, BATRA (1984) also observed that male fish had a higher intensity of infection than females and WILLIAM and JONES (1994) concluded that in general, male fish tend to be more heavily infected than females.

This study indicates that the presence of the nematode. Contracaecum. and acanthocephalan, P. kenyensis, did not seem to affect the body condition of O. leucostictus. This agrees with the findings of MALVESTUTO (1975) and MUCHIRI (1990). Elsewhere, TOMPKINS (1976) also reported that Contracaecum did not affect the body condition of O. leucostictus from Lake Turkana while MBAHINZIREKI (1984) likewise observed that unlike Eustrongyloides sp, Contracaecum seemed to be harmless to the Haplochromis spp. from Lake Victoria.

ACKNOWLEDGEMENTS

The authors wish to thank the German Academic Service Centre (DAAD) for providing funds for this work. The Director of Fisheries and his staff at Lake Naivasha Station provided technical assistance and laboratory space and we wish to thank them sincerely.

REFERENCES

- Aloo, P.A., 1995. Ecological studies fo helminth parasites of fish in Lake Naivasha, Kenya PhD. thesis Kenyatta University, Kenya. 137 p.
- Amin, O.M., 1985. The relationship between the size of some salmonid fishes and the intensity their acanthocephalan infections. Canadian Journal Zoology 63:924-927
- Batra, V., 1984. Prevalence of helminth parasites in three species of Cichilds from a man-made lake in Zambia. Zoological Journal of Linnean Society 82:319-333
- Dogiel, V.A., Petrushevski, G.K. and Yu I. Polyanski, 1958, Parasitology of Fishes. Oliver and Boyd. Edinburgh and London. 384 p.
- Fryer, G. and T.D. Iles, 1972. The Cichlid Fishes of the Great Lakes of Africa - Their Biology and Evolution. Oliver and Boyd, Edinbury. 641 p.
- Malvestuto, S., 1975. Contracaecumosis of Oreochromis leucostictus in Lake Naivasha. Kenya. M. Sc. thesis University of Nairobi, Kenya. 65 p.
- Malvestuto, S. and A. Ogambo-Ongoma, 1978. Observation on the infection of *Oreochromis* leucostictus (Pisces: Cichlidae) in Lake Naivasha, Journal of Parasitology 64: 383-384.
- Margolis, L., G. W. Esch, J.C. Holmes, A. M. Kuris and G.A. Schad, 1982. The use of ecological terms in Parasitology (Report in Ad hoc Committee of the American Society of Parasitologists). Journal of Parasitology 68 (1): 131-133.
- Mashego, N., 1989. A new species of Acanthosentis Veroa and Datta 1929 (Acanthocephala: Quadrigyridae) from Barbus neefi in South Africa. Annals of the Transvaal Museum 34: 545 - 549.
- Mbahinzireki, G.B.A., 1984. Parasite fauna of Haplochromis spp. (Pisces: Cichlidae) from Mwanza Gulf of Lake Victoria. M. Sc. thesis University of Dar-es-Salaam, 157 p.
- Muchiri, S. M., 1990. The feeding ecology of tilapia species and the fishery of Lake Naivasha, Kenya, PhD. thesis. University of Leicester. U.K. 233 p.

- 16 P.A. ALOO, R.O. OKELO AND M.J. NTIBA
- Paling, J.E., 1965. The population dynamics of the monogenean gill parasite *Disocotyle sagittata* Leuckart on the Windermere trout, *Salmo trutta Parasitology* 55: 667-694.
- Paperna, I., 1980. Parasites, Infections and Diseases of Fishes of East Africa. CIFA Technical Paper No. 7. FAO publication. Rome, Italy. 215 p.
- Paperna, I., 1991. Diseases caused by parasites in the Aquacualture of warm water fish. Annual Review of Fish Diseases. P. 155-194.
- Scholtz, T., 1986. Observation on the ecology of five species of intestinal helminths in the Perch (Perca fluviatilis) from the Macha lake fish pond system, Czechoslovakia. Vest. Cs. Spolec zool. 50:300-320.
- Tompkins, J.A., 1976. The effect of parasites on the condition of *Tilapia nilotica* from Lake Baringo, Kenya. Supplement to Lake Baringo Research Report. General Survey 1975 1976. Report to the Kenya Government. 91 p.
- William H. and A. Jones, 1994. *Parasitic worms of fish*. Taylor and Francis, London, U.K. 593 pp.